

# Effects of *hepA*, *rfaY*, and *cspC* Genes on Benzoate Tolerance and Antibiotic Resistance in Benzoate Evolved *Escherichia coli*

Morgan L. Engmann '20 and Joan L. Slonczewski

Department of Biology - Kenyon College - Summer Science 2017

## Abstract

*Escherichia coli* underwent evolution for 2000 generations in potassium benzoate conditions that increased in concentration over time. Strains were isolated from this experiment and testing them using growth curves revealed that strains had begun to show a resistance to benzoate yet began to be more sensitive to antibiotics (Creamer et al., 2016). After sequencing their genomes, many mutations were found that could be causing this phenotype. I focused on the mutations in the genes *hepA*, *rfaY*, and *cspC*. These genes were knocked out in the original background strain, W3110, and tested in several environments to observe growth differences. Growth curves revealed that the *hepA* knockout showed decreased growth in benzoate conditions, the *rfaY* knockout showed decreased growth in chloramphenicol (CHL), and the *cspC* knockout showed increased growth in benzoate conditions. For *hepA* and *rfaY*, these results raise more questions than they answer. Why would the bacteria select for mutations that cause decreased growth? The mutation in *hepA* is a late one, therefore it could be a corrective mutation, however results are not clear. With further testing, the genes responsible for the phenotype seen in the evolution experiment will be found and will hopefully explain why the bacteria selected to lose antibiotic resistance and if there is a possible tradeoff for benzoate tolerance.

## Introduction

For 2000 generations, *E. coli* underwent evolution in the presence of increasing concentrations of benzoate by the Slonczewski lab at Kenyon College. This environment created an acid stress that the bacteria had to grow and adapt in. Strains collected from this experiment have been phenotyped as having increased fitness in 15mM benzoate and having a lowered resistance to several antibiotics such as chloramphenicol (CHL) (Creamer et al., 2016). After these strains had been sequenced and compared to the wild type (D13) many mutations were found in the evolved generations. A few of these mutations included those in the genes of *hepA*, *rfaY*, and the intergenic region near *cspC*. These mutations were selected during evolution, therefore most are thought to have positive effects on the growth of the bacteria in the benzoate environment. For this reason, an important question has been asked of this experiment: why are the bacteria selecting for mutations that are causing decreased growth in antibiotics?

In order to begin to answer this question, mutations in genes within the evolved strains must be studied to determine their effect on growth. For my project, I chose to study the effects of many of these genes and I have found that growth in certain environments differs when a particular gene is knocked out, or deleted. The three genes I have found results with include *hepA*, *rfaY*, and *cspC*. The *hepA* gene is known to produce an RNA polymerase-associated protein and its knockout shows increased sensitivity to UV light (Muzzin et al., 1998). The gene *rfaY* (*waaY*) produces a protein involved in the making of the cell membrane and mutations in this gene can result in a "leaky" outer membrane (Ractz, 1996). *rfaY* mutants have also been seen to be more sensitive to substances such as menadione and plumbagin however they were more resistant to paraquat (Lee et al., 2009). The *cspC* gene codes for a cold-shock protein, and creating a knockout of this gene has been shown to confer an overall gain of fitness in normal broth cultures (Rath and Jawali, 2006).

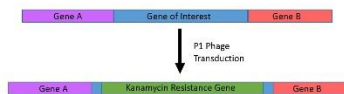


Figure 1: P1 phage transduction consist of replacing the gene of interest with a kanamycin resistance gene, creating a knockout. The use of the resistance gene allows for selection of transduced cells versus the original recipient cells.

## Mutations in *hepA*, *rfaY*, and *cspC*

Strain	A5-1	A5-4	A5-5	A1-1	C3-3	C3-4	C3-5	Mutation	Annotation	Gene
62,682								-G	coding (583-2907 nt)	<i>hepA</i> --
1,908,877								C→A	intergenic (-103-+43)	<i>rfaA</i> -- / -- <i>cspC</i>
3,840,032								Δ4 : insR 155(-)+4 bp : Δ4	coding (585 -586-699 nt)	<i>rfaY</i>

Table 1: Mutations in *hepA*, *cspC*, and *rfaY* occur in strains of benzoate evolved *E. coli*. The whole genome sequences were matched to the reference genome of *E. coli* K-12 W3110. Mutations were found using *breseq* analysis.

## *hepA* knockout showed decreased growth in benzoate compared to the background strain

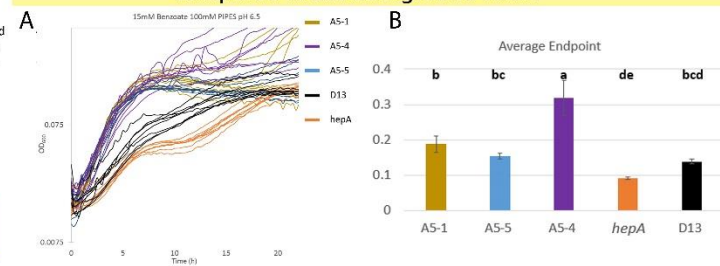


Figure 2. A+B: The *hepA* knockout strain was grown in 15mM potassium benzoate 100mM PIPES pH 6.5 along with benzoate evolved strains and its background strain (D13). Endpoint data from t=16h was analyzed (ANOVA w/Tukey, F=26.94 p=2e-16, n=8 for each strain).

## *rfaY* knockout showed decreased growth in CHL compared to the background strain

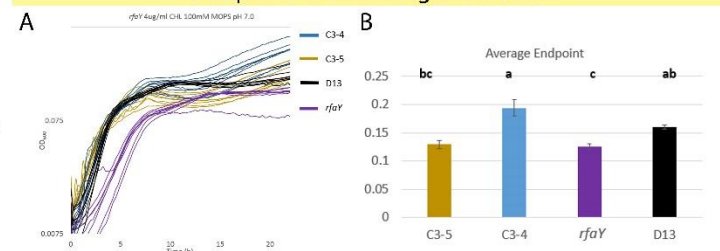


Figure 3. A+B: The *rfaY* knockout strain was grown in 4ug/ml CHL 100mM MOPS 5mM potassium benzoate at pH 7.0 along with its background strain (D13) and benzoate evolved strains. Endpoint data from t=16h was analyzed (ANOVA w/Tukey, F=11.08 p=1.06e-08, n=6 for each strain)

## *cspC* knockout showed increased growth in benzoate compared to the background strain

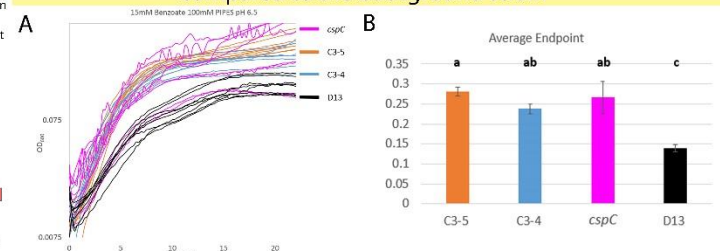


Figure 4. A+B: The *cspC* knockout strain was grown in 15mM potassium benzoate 100mM PIPES buffer at pH 6.5 along with benzoate evolved strains and its background strain (D13). Endpoint data from t=16h was analyzed (ANOVA w/Tukey, F=76.16, p=2e-16, n=8 for each strain)

## Methods

**Construction of Knockouts:** The targeted gene was interrupted by a kanamycin resistance gene through P1 phage transduction. See figure 1.

**Colony PCR:** To check if the transduction worked, PCR was used to identify changes in length of the targeted region. Primers were designed using NCBI Blast

**Growth Curves:** Using a spectramax spectrophotometer, the growth of the *E. coli* was measured using OD values that were measured every 15 minutes for 22 hours. The growth curves were read in sterile 96 well plates that contained 200ul of LB based growth medium inoculated with 1ul of *E. coli*.

## Conclusions

~The *hepA* knockout shows decreased growth rate yet same endpoint in 15mM benzoate compared to D13 (Figure 2). The mutation in the evolved strain may be a corrective mutation.

~The *rfaY* knockout shows decreased growth in 4ug/ml of CHL in both growth rate and endpoint (Figure 3). The growth is less than that of the evolved strains. The *rfaY* knockout does not show sensitivity to benzoate, salicylate, or ampicillin.

~The *cspC* knockout shows increased growth rate and endpoint in 15mM benzoate compared to D13 (Figure 4). The knockout grew similarly to C3-5 and C3-4 despite the mutation only being present in C3-5.

## Acknowledgements

I would like to thank Dr. Slonczewski and all of the members of her lab for their support, guidance, and aide in this project. Special thanks to those members of the lab that were also here over the summer including Trudy Wrona, Haofan Li, Carter Brzezinski, Preston Basting, and Katie Bischof as well as our high school helpers Mary Harris, Ellie Broeren, and Chase Holdener for their help and mentorship throughout this project. I would also like to thank Karina Kunka for compiling the mutation chart. This work was funded by the Kenyon Summer Science Program.

## References

- Creamer K, Ditmars F, Basting P, Kunka K, Hamdallah I, Bush S, Scott Z, He A, Penix S, Gonzales A, Eder E, Camperchioni D, Berndt A, Clark M, Rouhier K, Slonczewski J. 2017. Benzoate and Salicylate Tolerant Strains of *Escherichia coli* K-12 Lose Antibiotic Resistance during Laboratory Evolution. *Applied and Environmental Microbiology*, 83 (2), pp. 1-19.
- Lee J, Lee K, Yeo W, Park S, Roe J. 2009. SoxRS-Mediated Lipopolysaccharide Modification Enhances Resistance against Multiple Drugs in *Escheria coli*. *Journal of Bacteriology*, 2009, pp. 4441-4450.
- Muzzin O, Campbell E, Xia L, Severinova E, Darst S, Severinova K. 1998. Disruption of *Escherichia coli* *hepA*, an RNA Polymerase-associated Protein, Causes UV Sensitivity. *Journal of Biological Chemistry*, 273, pp. 15157-15161.
- Ractz, C.R.H. 1996. Bacterial lipopolysaccharides: A remarkable family of bioactive macroamphiphiles. In *Escherichia coli and Salmonella. Cellular and Molecular Biology*. Washington DC: American Society for Microbiology Press, pp. 1035-1063.
- Rath D, Jawali N. 2006. Loss of Expression of *cspC*, a Cold Shock Family Gene, Confers a Gain of Fitness in *Escherichia coli* K-12 Strains. *Journal of Bacteriology*, 2006, pp. 6780-6785.