



# Characterization of a zebrafish peripheral myelin mutant

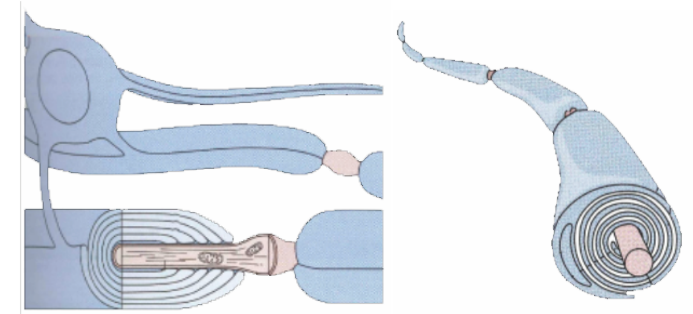


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## Zebrafish provide an ideal system for studying the development of myelinating glia

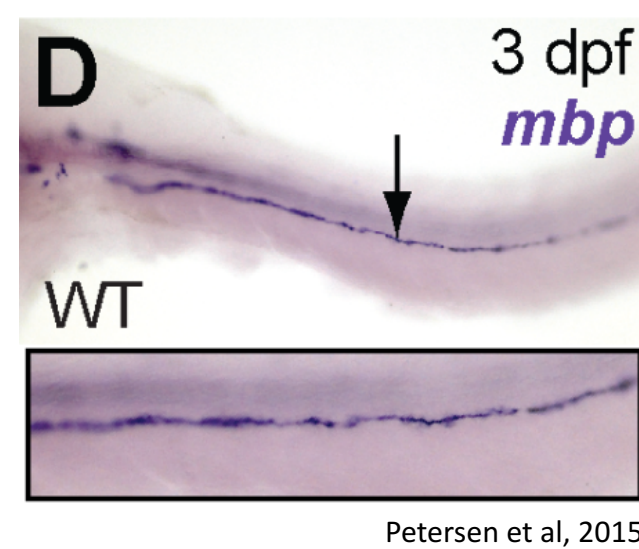
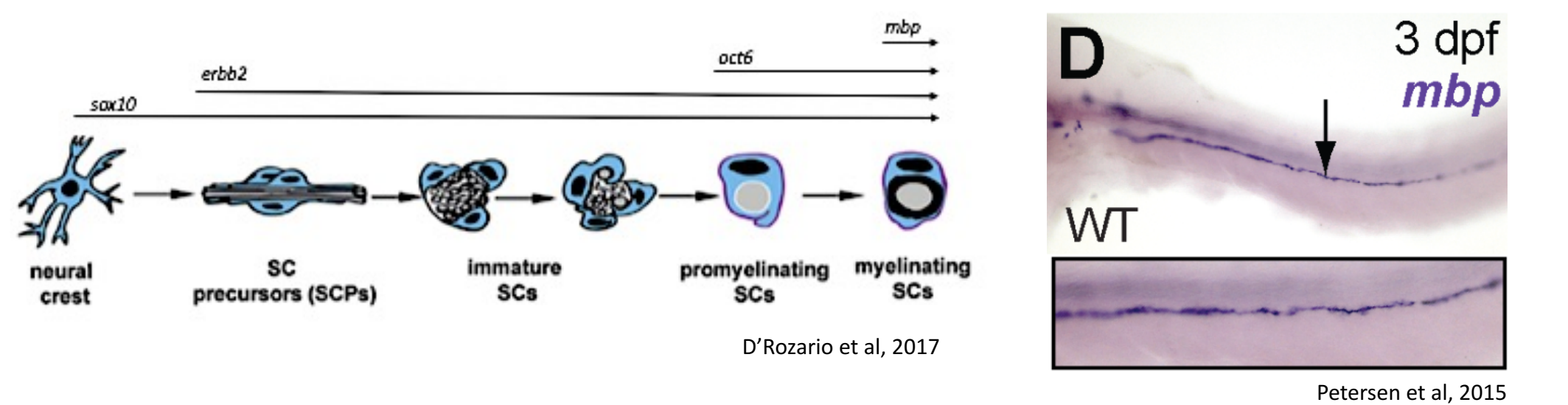
Myelin insulates axons and allows for efficient electrical signaling

Myelin is produced by Schwann cells in the PNS and oligodendrocytes in the CNS



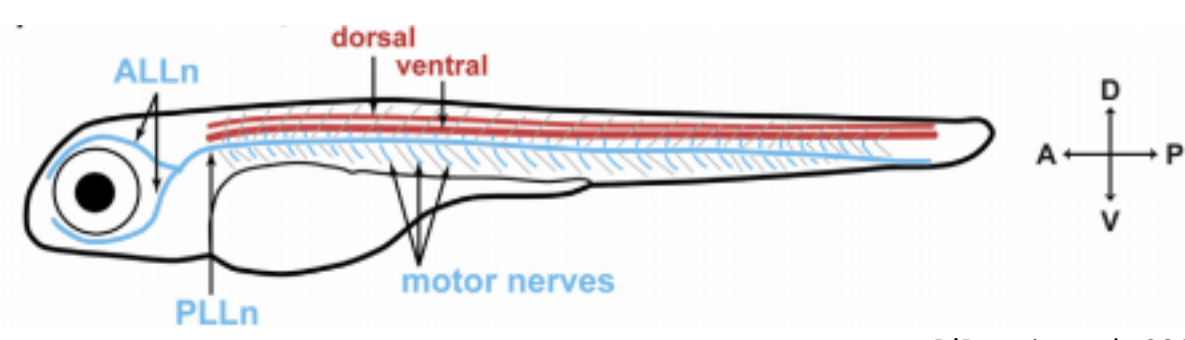
Kandel et al., 2006

Each stage of Schwann cell development can be defined by key developmental markers



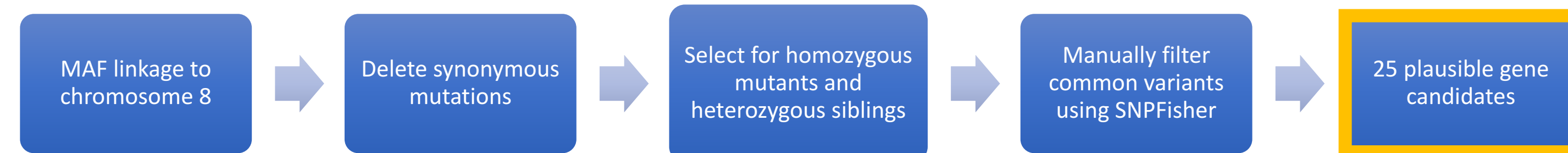
Petersen et al., 2015

Mechanisms of myelination is conserved in jawed, vertebrate organisms



D'Rozario et al., 2017

## Whole-genome sequencing analysis pipeline to identify affected gene(s) in *st144*



## Whole Genome Sequencing of *st144* shows linkage to chromosome 8

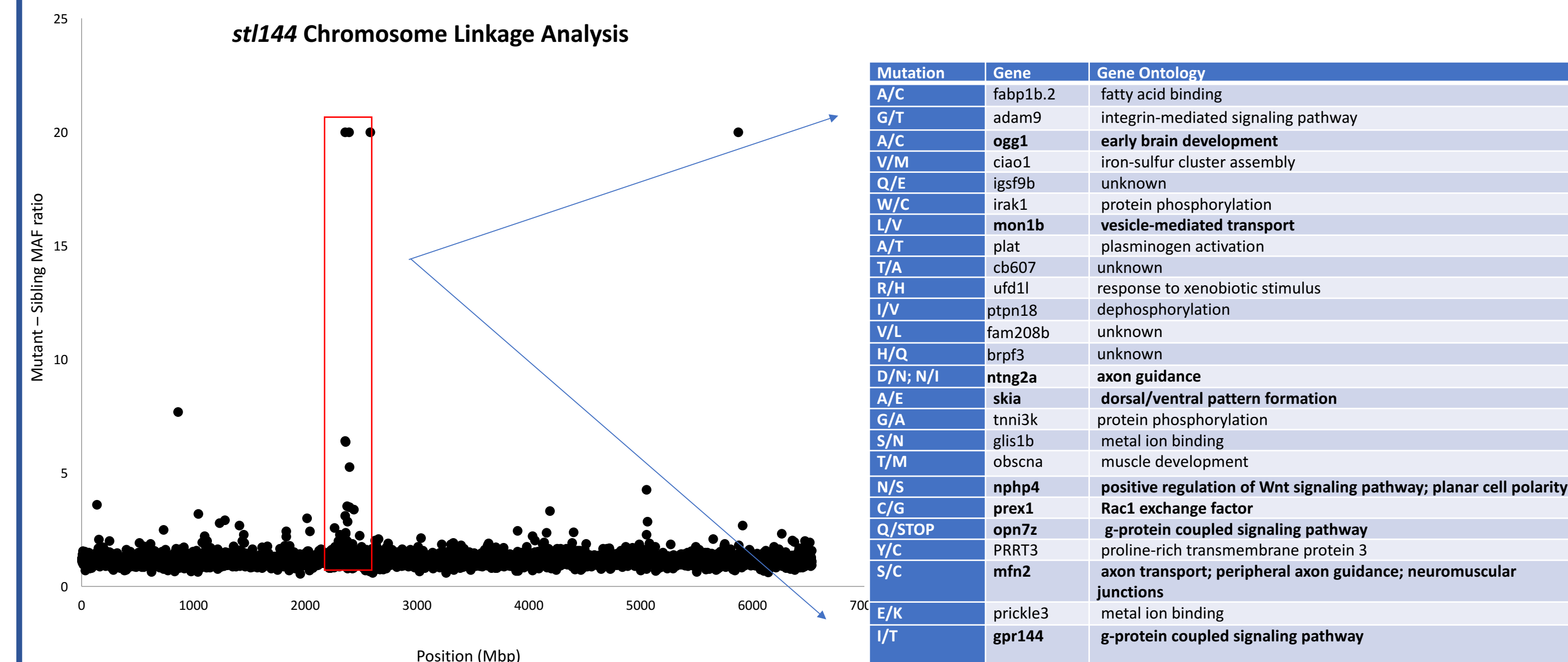
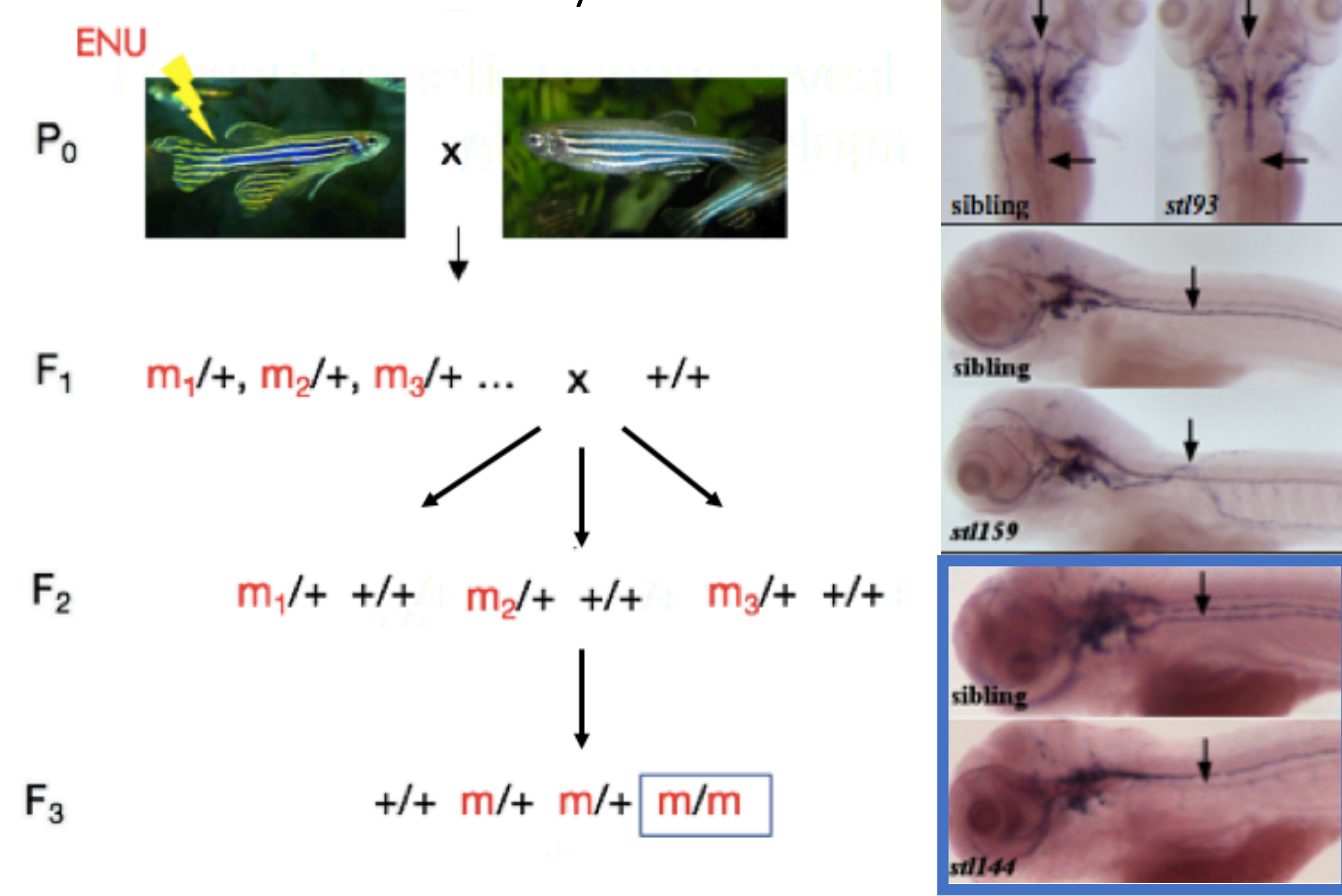


Figure 1. Whole genome sequencing of mutants and siblings shows linkage to chromosome 8. Regions of linkage were located using mutant-sibling MAF ratio mapping. 22 candidate genes were identified.

## Forward genetic screens uncover genes that regulate myelination in vertebrate systems

A screen conducted at Washington University, St. Louis revealed three novel mutants with reduced myelination.



## *st144* homozygous mutants have reduced PNS myelination

- Where is the causative lesion? What gene does it affect?
- At what point in development is this gene necessary?
- What are the developmental effects of disrupting the gene?
  - Reduced number of Schwann cells
  - Reduced wrapping by Schwann cells
  - Reduced number of axons present for wrapping

## Identifying Carriers of *st144* By Cross

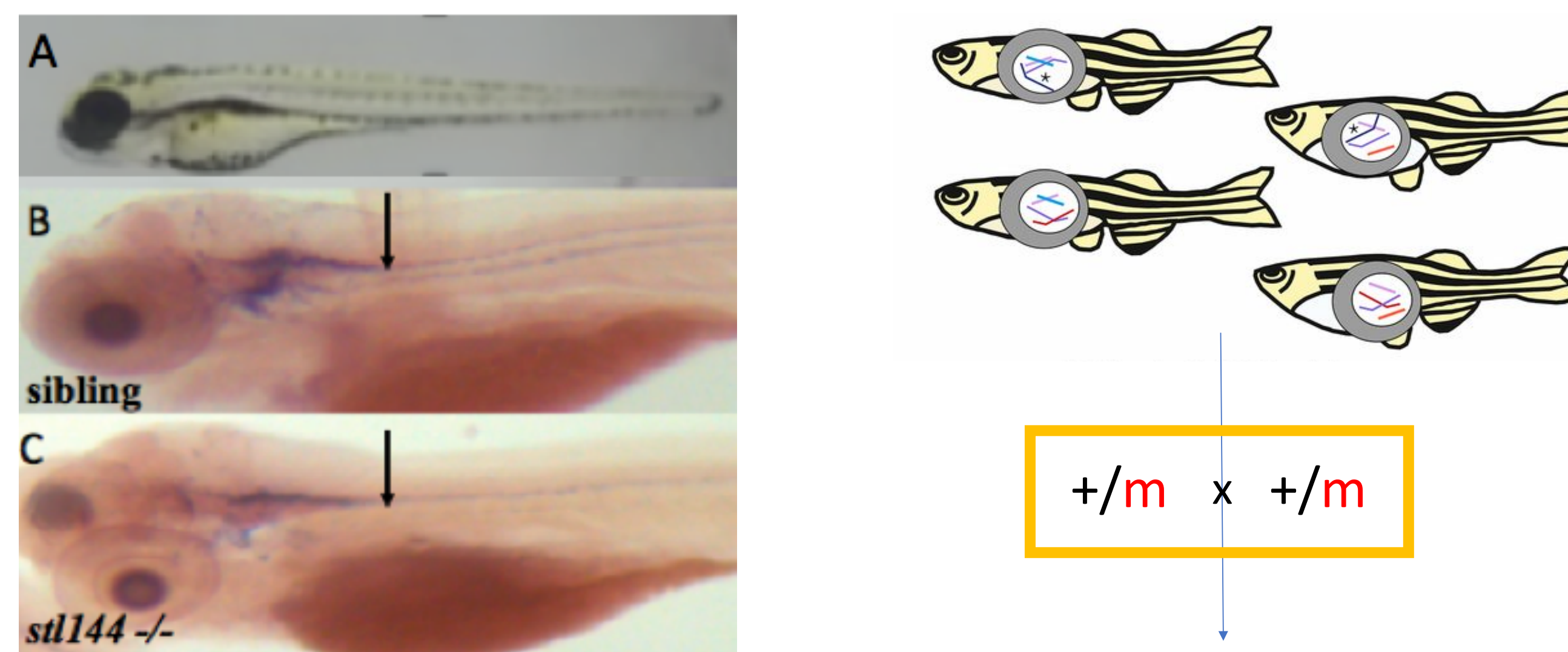


Figure 2. Reduced expression of mbp reveals *st144* homozygous mutants in the progeny of a random in-cross (n=6/23). (A) Live progeny showing no visible deformities. (B) *st144* sibling with wild-type expression of mbp along the pLLn (arrow). (C) *st144* -/- mutant with reduced expression of mbp along the pLLn.

## Acknowledgments

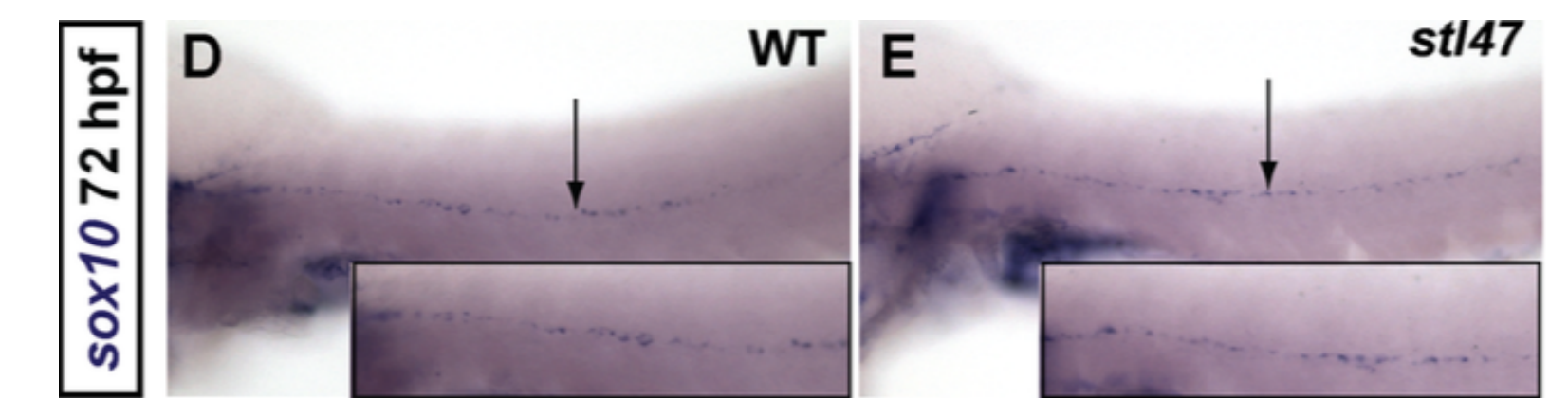
I would like to thank Dr. Sarah Petersen for her guidance and mentorship, as well as the Petersen lab group for their continued support. I thank Becky Gallagher and the Kenyon animal care staff for their excellent maintenance of the Higley Aquaculture Facility.

I thank Nicolas Sanchez in the Monk lab for his troubleshooting of the whole genome sequencing data. Finally, I thank the Monk lab at Washington University, St. Louis for conducting original the forward genetic screen that identified the aforementioned myelin mutants.

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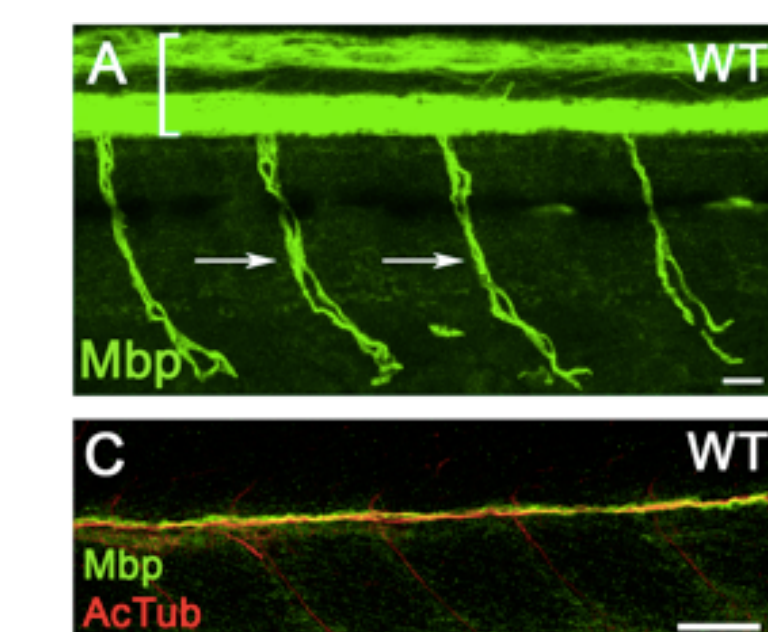
## Future Directions

- Are neural crest cells present and migrating properly?
  - sox10* marks neural crest cells and can reveal if early development is affected by the mutation.



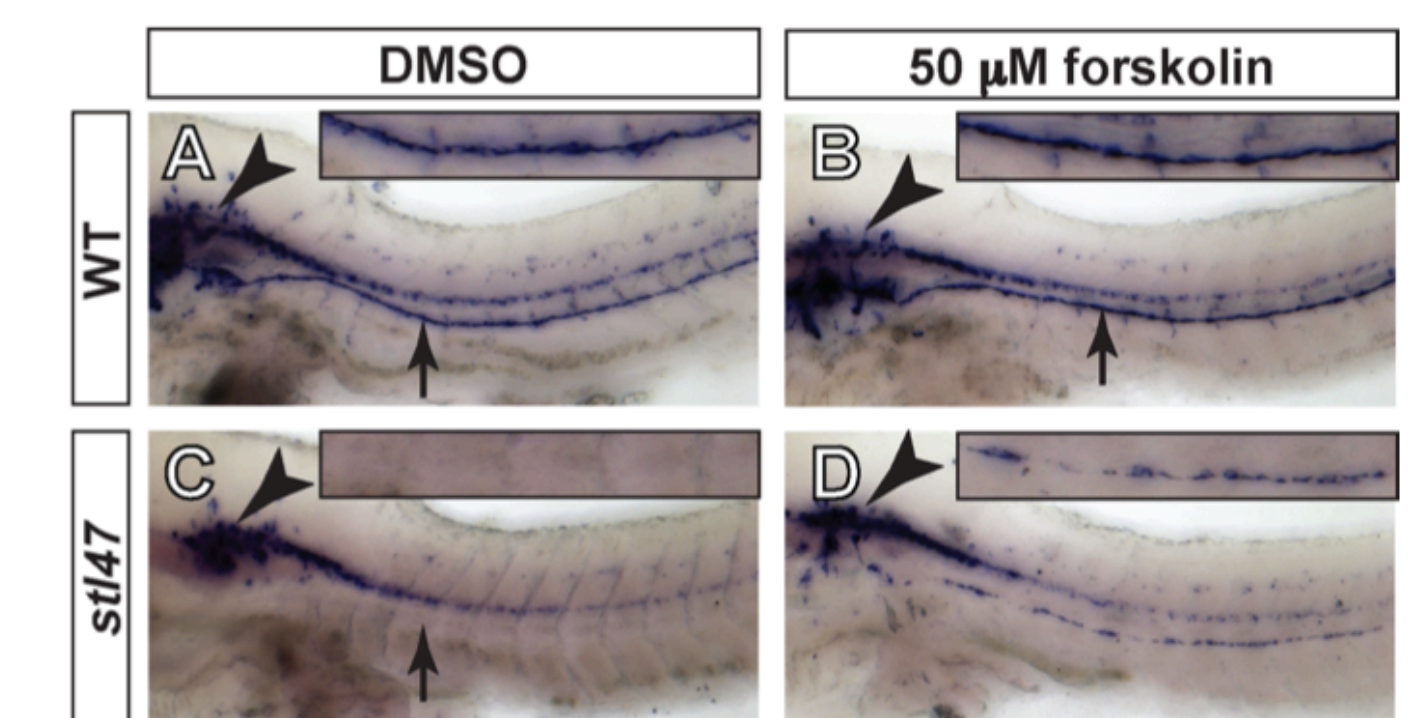
Petersen et al., 2015

- Are axons present and migrating normally?
  - Acetylated microtubules mark PNS axons and can be used to determine if axon development is affected in *st144* mutants.



Monk et al., 2009

- Is the *st144* mutation affecting a GPCR, upstream of cAMP?
  - Rescuing *st144* mutants with forskolin, a cAMP elevator, may reveal the genetic pathway affected by *st144*.



Petersen et al., 2015

Molecular characterization of *st144* mutants may shed light on which candidate gene is most likely responsible for the reduction in PNS myelination.

## References

Monk K. R. et al. . A G protein-coupled receptor is essential for Schwann cells to initiate myelination. *Science* 325, 1402–1405 (2009)

Monk K.R., Oshima K., Jors S., Heller S., Talbot W.S. Gpr126 is essential for peripheral nerve development and myelination in mammals. *Development*. 2011;138:2673–2680. doi: 10.1242/dev.062224.

Petersen SC, et al. The adhesion GPCR GPR126 has distinct, domain-dependent functions in Schwann cell development mediated by interaction with laminin-211. *Neuron*. 2015;85(4):755–769. doi: 10.1016/j.neuron.2014.12.057.

D'Rozario M, Monk KR, Petersen SC. Analysis of Myelinated Axon Formation in Zebrafish. *Methods in Cell Biology: The Zebrafish*, 4th edition. Volume 138, Pages 383-414 (2017).